Chikungunya: Molecular Aspects, Clinical Outcomes and Pathogenesis

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ABSTRACT

The alarming worldwide emergence of the chikungunya virus began in the last decade. Since the first autochthonous transmission in Mexico in November of 2014, the virus has spread throughout the country, resulting in multiple outbreaks. This virus produces an acute and self-limiting disease characterized by fever, polyarthralgia, myalgia, exanthema, and general malaise. It is transmitted to humans by the bite of Aedes aegypti and A. albopictus mosquitoes. The fact that the clinical presentation is similar to that produced by other arboviruses complicates its clinical diagnosis. The chronic stage of the disease can cause severe consequences lasting months or years, from local arthralgia to rheumatoid arthritis. In this review, we emphasize the public health threat posed by this highly disabling emerging disease, the clinical outcomes, and its possible physiopathological process. We outline the diagnosis and the impact that this virus has had in Mexico since its introduction.

Key words: Chikungunya. Pathogenesis. Molecular aspects.

INTRODUCTION

Chikungunya virus (CHIKV) is an emerging arbovirus in the Americas that affects humans, causing chikungunya fever. The virus was initially isolated from human serum and infected mosquitoes during an epidemic in Tanzania in 1953. In 2004, CHIKV reappeared on the coast of Kenya and spread to the Comoro islands, where 5,000 cases were reported1. In 2005-2006, the virus disseminated to other islands of the Indian Ocean, mainly La Réunion where it is estimated that 300,000 people out of a population of 785,000 were infected, leading to 237 deaths2,3. The epidemic reached the Asian continent in 2006 when large outbreaks in India affected an estimated 1.5 million individuals4.

On December 6, 2013, the Pan American Health Organization (PAHO) confirmed the autochthonous transmission of CHIKV in the Americas, detecting cases of the infection in the island of San Martín5,6. The entry of the virus to this continent has led to its spread to different areas of the Americas, mainly countries in the
For CHIKV in the Americas and its association with poor control measures has been published\(^\text{13}\).

**GENERAL ASPECTS AND VIRUS REPLICATION**

The virus belongs to the Alphavirus genus in the Togaviridae family, and is made up of an icosahedral capsid and a membrane envelope. The size of the virion varies from 60 to 70 nm. Its genome consists of a single-stranded, positive-sense RNA molecule of approximately 12 Kb. The genome consists of two open reading frames (ORF) that codify for nine proteins: five structural proteins (SP), including the capsid (C), the membrane (6k), and three envelope proteins (E1, E2, E3); and four non-structural proteins (NSP), including NS1, NS2, NS3 and NS4\(^\text{14}\).

In Mexico, in late 2014 during an outbreak of febrile illness in Chiapas, 79% of analyzed samples tested positive for CHIKV\(^\text{9}\) and *A. aegypti* was identified as its primary vector\(^\text{10}\). In 2015, 11,577 cases were confirmed, with four deaths, mostly in the south-central states of Mexico (DGE/InDRE, 2016; OPS/OMS, 2016). An extensive sequence analysis of autochthonous Mexican virus isolates showed that these belonged to the Asian genotype\(^\text{11}\). In Yucatan, Mexico, CHIKV from the Asian lineage was also isolated from febrile individuals, one case in coinfection with DENV-1\(^\text{12}\). An extensive discussion on the pattern of transmission for CHIKV in the Americas and its association with poor control measures has been published\(^\text{13}\).
There are three genotypes of CHIKV, named after the geographic region in which they were first detected: the West African genotype, the Asian genotype, and the East/Central/South African genotype (ECSA). To date, no serotypes of CHIKV have been found, meaning that all genotypes are antigenically similar.15,16

The name of the virus and disease comes from the Bantu language of the Makonde people, meaning “that which bends up”, referring to the curved posture of people with the disease, resulting from the painful arthralgia produced by an infection with CHIKV.17,18 CHIKV is capable of propagating in an ample variety of the infected cells of hosts, which include humans, monkeys, birds, cattle, and rodents. In humans, the surface glycoprotein E219 of the virion interacts with susceptible host cells that present receptors to this protein on their surface.

One of the proposed receptors for this virus is prohibitin-1 (PHB-1). However, not much is known about the participation of this protein in viral pathogenesis.20 The PHB-1 is a protein of 32 kDa involved in the regulation of proliferation and apoptosis, among other functions. This protein is normally located in the mitochondria and its expression is affected by high levels of interferon (IFN)-γ and tumor necrosis factor (TNF)-α. It is also known that the exogenous expression of PHB-1 reduces basal autophagy as well as that induced by TNF-α.21

After the interaction between the viral particle and the host-cell receptor, the former is internalized in the cell through endocytosis. Inside the endosome, there is a reduction in pH due to the pumping of H+ ions, which leads to a conformational reorganization of the E1-E2 heterodimer. In the glycoprotein E1, dominion II is exposed and fusion is carried out between the membrane of the endosome and that of the viral particle, thus releasing the nucleocapsid of the viral genome.13 The interaction of the largest RNA subunit 60S with the proteins of the nucleocapsid signals its decoupling, thus releasing the genome. This is a highly conserved mechanism among Alphavirus.14

Once the viral genome (49S) is released in the host cell, the translation of the first ORF is carried out, this codifying for the synthesis of non-structural proteins (NSP1, NSP2, NSP3, NSP4) by means of the production of a polyprotein named P1234. The latter is processed proteolytically, generating the complex of viral replication (Fig. 2). NSP1 participates in the synthesis of the complementary chain of viral RNA as well as in the methylation and binding of CAP to the 5’ extreme of the viral genome. NSP2 presents activity of RNA helicase, RNA triphosphate and protease, and inhibits the transcription of cellular mRNAs. NSP3 participates in the production of the negative chain of viral RNA. NSP4 is the polymerase RNA dependent on DNA.22

After the first ORF is translated, the complementary chain of viral RNA (negative chain) is produced. This serves as a template for the transcription of the second ORF known as subgenomic RNA 26S. The translation of this ORF gives rise to the generation of the structural proteins, C, E3, E2, 6k and E1. The C and 6k proteins are accumulated in the cellular cytoplasm to form new nucleocapsids, which are assembled with a copy of the viral genome by means of proteolytic cuts.

Proteins E3, E2, and E1 undergo post-translation processing, being glycosylated in the endoplasmic reticulum. They are then sent to the Golgi apparatus to be later transported in vesicles to the cell membrane. When the nucleocapsid interacts with the accumulated glycoproteins in the cell membrane, the viral particles undergo a maturation process by acquiring the membrane envelope, and then finally are released by means of exocytosis.17 It is important to mention that whereas the synthesis and translation of subgenomic 26CC RNS remains constant, the synthesis of the NSPs decreases and the transcription of genomic RNA 49S increases to the extent that the infection progresses inside the cell.14,19

Once a mosquito infected with CHIKV bites a healthy individual, the virus first multiplies in the fibroblasts located in the epithelium, and then, through infected macrophages, it disseminates to the lymph nodes. Additionally, the virus disseminates to the liver and joints through the blood flow that transports the virions produced.17 The virus continues to propagate itself, evidenced by a viral load of up to 10^8 PFU/ml (viremia phase) that has been detected at these sites.22 Later, it disseminates to the entire organism, thus initiating the signs and symptoms of the disease. The incubation period varies from 2-12 days.18 The active infection only occurs in cells permissive to CHIKV. In mice, these cells primarily consist of muscle, joints, and skin fibroblasts. However, they have also been found in epithelial and endothelial cells of different organs, such as the liver, spleen, and brain.17
CHIKV induces a self-limiting disease, as it is eventually eliminated from the organism. The mechanisms of the innate immune response, mainly the production of IFN-1α/β, and later those of adaptive immunity, the IgM and IgG antibodies, circumscribe the infection and finally control and eliminate it from the organism 23,24.

TRANSMISSION OF CHIKV

CHIKV is transmitted to humans through a bite from a female Aedes mosquito, especially two species17, A. aegypti and A. albopictus. The former is considered the primary vector in the transmission of the disease since it is commonly associated with human habitat. However, during the outbreak of this virus on La Réunion Island in 2006, as well as in Africa and India, A. albopictus was also identified as an efficient vector, particularly from the ECSA genotype. This event was attributed to adaptive mutations in the genome of CHIKV within the codifying genes for two SPs, E1 and E216, mainly involving the mutation of A226V in the E1 protein. This change favors replication within the A. albopictus vector and the efficiency of the interaction of the virus with the host cell receptor15.

The virus circulates in nature through two cycles of transmission: the sylvatic (enzootic) cycle and the urban (epidemic) cycle, which are found in distinct geographic areas24. In Africa, CHIKV is maintained through the sylvatic cycle, in which non-human primates are the primary reservoirs. This situation leads to frequent outbreaks in urban areas15. Typically, these outbreaks coincide with the season of intense rain and the corresponding increase in mosquito population density. The species of mosquitoes that transmit CHIKV in Africa have been identified as A. furcifer, A. taylori, A. vittatus, A. fulgens, A. luteocephalus, A. dalzieli, A. vigilax, A. camptorhynites and A. africanus. Culex annulirostris and Mansonia uniformis are also considered competent vectors25.
As a consequence of population movements from endemic zones of Africa (e.g. Kenya and Comoro islands) to territory where the vector but not the virus previously existed, CHIKV has become an emerging virus. During the years 2005 and 2006, there were multiple outbreaks in India and the islands of the Indian Ocean, including La Réunion Island, in urban cycles of viral transmission. In these cycles, humans in urban zones constitute the main host and the virus disperses successfully by means of human-mosquito-human transmission. The species that participate in this transmission are *A. aegypti* and *A. albopictus*, as previously mentioned, leading to massive outbreaks.

Once the mosquito bites an infected host, the virus arrives to the intestinal tract of the insect, multiplies in the vector’s cells, and disseminates to the salivary glands. Then, the mosquito can transmit the virus in each feeding for the rest of its lifetime. To date, there has been no consistent evidence of transovarial transmission in the vectors.

### CLINICAL PRESENTATIONS

With a mosquito bite, viral particles are deposited in the skin, from where they eventually reach the lymph nodes and then the blood flow. They are then distributed to target organs that include joints, muscle, and skin (Fig. 3). With less frequency, the virus can affect the liver or cause encephalopathy, encephalitis, myocarditis, and heart blockage. Although 5-15% of individuals suffer from asymptomatic infection, the rest generally develop symptoms that progress from the acute to the chronic phase.

**Acute phase**

The acute phase lasts 3-10 days and is characterized by the abrupt presentation of fever (≥ 39 °C). Other symptoms follow during the next few days. Joint pain is reported in 87-98% of the cases. These two symptoms are the most characteristic of this disease.

Joint pain exists in more than one joint and is usually bilateral (symmetrical), occurring mainly in the peripheral joints (knees, ankles, hands, wrists). There is myalgia in 46-72% of the cases, affecting arms, thighs, and calves. This clinical picture is called chikungunya fever (CHIKF) and tends to be limiting and even disabling in relation to the normal physical activity of individuals.

The exanthema that appears in 40-50% of cases is of the petechial or maculopapular type, mainly affecting the limbs, trunk, and face, and can cause pruritus. The lesions are transitory and generally appear 2-5 days after disease onset. Other symptoms that are less common are diarrhea, vomiting, edema on limbs, bleeding, otitis, and ocular disease (especially anterior uveitis). Some of the more severe manifestations, which are infrequently found, include neurological diseases such as meningoencephalitis, the Guillain-Barré syndrome, myocarditis, and multiorgan failure. The latter can be fatal, particularly in neonates and older adults with comorbidity. Laboratory findings from persons in the acute stage of the disease have evidenced lymphopenia (500-1000 cells/mm³) and moderate thrombocytopenia (100,000-150,000 cells/mm³). Other uncommon abnormalities include leukopenia, elevation of hepatic enzyme levels, anemia, elevated creatinine level, and hypocalcemia.

In neonates, CHIKF can be accompanied by convulsions, peripheral cyanosis, podalic edema, and epithelial vesicular lesions that eventually dry and scale. This disease is usually considered benign in children. However, neurological manifestations have been reported that include febrile convulsions, meningeal syndrome, acute encephalopathy, diplopia, aphasia, acute disseminated encephalomyelitis, and encephalitis. The development of chronic arthralgia and exacerbation of underlying medical conditions is unusual.

**Chronic phase**

After the diverse outbreaks and epidemics of chikungunya occurred in different regions of Africa, Asia and Europe, follow-up studies generated sufficient evidence to indicate that the infection with this virus can induce chronic rheumatic diseases. In adults, these conditions can last months or even years after the infection, while they are less common in children.

The musculoskeletal disorders include arthralgia, inflammatory arthritis, polyarthralgia, tenosynovitis, enthesitis, and exacerbation of existing rheumatic disease. Other less common symptoms include neuropathy, cerebrovascular disorder, neurosensory deficiency, burning mouth syndrome, paresthesia, cubital
tunnel syndrome, gastrointestinal disorders, exan-
thena, pruritus, bursitis, and synovitis. Infection with CHIKV has been associated with destruc-
tive rheumatoid arthritis, which is similar to rheumatoid
arthritis and induces a similar inflammatory response,
although the laboratory tests are negative for the rheu-
matoid factor and the antibodies for cyclic citrullinated
peptides. Radiographic studies of some of these individuals show bone erosion, swollen joints and
synovitis. The symptoms can be recurrent or chronic
and can affect multiple joints (mainly those previ-
ously injured), which significantly diminishes the quality
of life of the patient both physically (regarding function,
pain and general health) and psychological.

The development of arthralgia may be due to continu-
ous inflammation in joints in response to viral antigens,
evidenced by the fact that viral RNA has been detected
in perivascular macrophages. Risk factors for the de-
velopment of chronic arthralgia identified to date
include advanced age, long duration of the acute phase
of CHIKF with more than six joints manifesting pain,
existing joint disease, and a delayed adaptive immune
response (mainly IgG neutralizing antibodies).

**Congenital infection**

Pregnant women with detectable viremia a few days
before childbirth can transmit CHIKV to the newborn,
which can result in a severe neonatal disease, gener-
ally encephalopathy followed by neurodisability. Other signs and symptoms observed in newborns are
convulsions, thrombocytopenia, hypotension, ventricu-
lar dysfunction, pericarditis, hyperechoic coronary ar-
teries, parenchymal hemorrhage, and cytotoxic ede-
ma. It has been demonstrated that maternal-fetal
transmission occurs due to contact of the product
with infected maternal blood during childbirth.

There is no specific antiviral drug treatment against
CHIKV. Nonetheless, symptomatic treatment includes
acetaminophen to relieve the fever and nonsteroid
anti-inflammatory drugs (e.g. paracetamol) for poly-
arthralgia.

**PATHOGENESIS OF CHIKUNGUNYA
VIRUS DISEASE**

*Alphavirus*, including CHIKV, cause severe forms of
diseases such as chronic and highly disabling arthral-
gia/arthritis. Chronic symptomatology is observed
with greater frequency in persons over 60 years of age that have very high viremia (> 10^10 PFU/ml) during
the acute phase of the disease. The infection is
initially very rapid and the virus is typically eliminated
5-7 days after onset of the fever. During the infection
there is a robust and rapid activation of dendritic, NK/
CD4+ and CD8+ cells. Persistently high levels of IL-12
are mainly found in individuals with chronic symp-
toms, mostly in persons with rheumatoid arthritis, as
inflammation provoked by CHIKV infection can cause
bone erosion and severe arthralgia.

The activity of osteoclasts increases, thus promoting
bone absorption. Additionally, the activity of osteo-
blasts is inhibited during infection, which leads to the
inhibition of bone formation. The differentiation of
osteoclasts from their precursors requires participation
of the inductor cytokine RANKL (receptor activator for
the nuclear factor kB ligand)\(^{60}\) and the interaction with its RANK receptor\(^{61}\). The activity of RANKL is inhibited by osteoprotegerin (OPG), a natural receptor that functions as a decoy for RANKL and blocks its interaction with RANK\(^{62}\) (Fig. 4). The concentrations of OPG and RANKL are maintained in a proportion that regulates the process of osteoclastogenesis\(^{63}\). When the concentration of RANKL rises, the production of osteoclasts increases, which leads to greater bone absorption.

Infections by Alphavirus share some common characteristics with other diseases such as rheumatoid arthritis\(^{64}\). Replication of CHIKV inside cells in biopsies of synovial tissues has been clearly demonstrated. Pulkka, et al. showed the direct infection of primary synoviocytes with CHIKV. These infected cells attract monocytes/macrophages and induce the formation of osteoclasts\(^{56}\). Primary cultures of osteoblasts have great susceptibility to infection with CHIKV, an infection that promotes the formation of osteoclasts and the loss of bone tissue\(^{65}\).

The innate immune system acts as the front line against an invasion by Alphavirus. There is increasing evidence of an immunopathogenesis due to the over-activation of the immune system by these viruses\(^{66}\). Apparently, the infiltration of macrophages to the joints determines the severity and persistence of the infection. The persistent infection of perivascular synovial macrophages by CHIKV and the constitutive infiltrate of CD14\(^+\) monocytes to the interior of the synovial cavity have been observed 18 months after the onset of symptoms in individuals infected with CHIKV\(^{54}\). During such an infection, a large number of cytokines, chemokines, and growth factors are stimulated in the plasma of these persons. Accordingly, a variety of interleukins have been detected: IL-1beta, IL-5, IL-6, IL-7, IL-10, IL-15 and IL-17. Ligands of chemokines found include CCL-3, CCL-4, CCL-2 (MCP-1) and CCL-5 (RANTES). Finally, granulocyte-macrophage colony-stimulating factor (GM-CSF) has been found\(^{67,68}\).

In synovial liquid of infected individuals, there are important levels of CCL-2, IL-16 and IL-8\(^{65}\), as well as elevated concentrations of RANKL, which could directly lead to bone loss\(^{69}\). These observations correlate with findings in animal models, such as mice and primates, in which it has been demonstrated that IL-6 promotes the formation and liberation of RANKL. Hence, a probable strategy for controlling the infection is the use of anti-IL-6 antibodies\(^{70}\).

In the infection by Alphavirus, an increase has been found in the activation of T CD4\(^+\) lymphocytes\(^{54,71}\), which differentiate to Th17 and infiltrate the synovial space, possibly promoting local inflammation\(^{72}\).

**FINAL REMARKS**

It is important to consider that diverse diseases have a clinical presentation similar to that observed with a CHIKV infection. There are a great number of clinical conditions that must be ruled out to confirm the diagnosis and these include: leptospirosis, malaria, infections with other alphaviruses, and other viral entities that cause exanthema.

Even though CHIKF is characterized by fever with polyarthralgia, these indicators are unspecific. Moreover, the infection can take place in a subclinical form or coexist with other infections like dengue\(^{65}\). Therefore, diseases that should be considered in a differential diagnosis vary in relation to the relevant epidemiological characteristics, such as place of residence, travel to endemic zones, and exposure to mosquito bites\(^{6}\).

In the differential diagnosis of CHIKV in Mexico, it is of great importance to consider DENV, due to a high similarity between the clinical manifestations of these two infections, as well as the high incidence of the latter agent in this country. Symptoms like myalgia, arthralgia, and exanthema are more associated with CHIKV, while thrombocytopenia may be more indicative of dengue\(^{40,67}\). Despite the broad similarity between both diseases, CHIKF has a more acute onset and lasts a shorter time; additionally, maculopapular exanthema is more frequent and joint pain is more intense/localized and highly incapacitating.

In the absence of a vaccine or antiviral drug on the market, vector control is the only currently available strategy for controlling CHIKV. The previous experience with DENV control efforts is not encouraging. The virus spread throughout the Americas was imminent due to inefficient vector control and the emergence of resistance to insecticides in mosquitoes. Nevertheless, it is essential to continue efforts to improve epidemiological surveillance of CHIKV and other viruses by strengthening the health services and facilities at different levels, mainly for primary care, to detect new cases of the virus in a timely manner. Moreover,
efforts to develop novel treatment strategies to fight this severe viral infection should continue.

REFERENCES